RESPONSE OF ANTIOXIDANT DEFENCES TO SELENIUM STRESS IN ALFALFA LEAVES

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Abstract

After three-month cultivation, the plants were treated with three Na_2SeO_3 concentrations of control, 100 and 900 μ M for 20, 40 and 60 days. Selenium (Se) induced a increase in SOD, CAT, APX activities and the contents of MDA, soluble protein but an decrease in POD activities and free proline contents in leaves. The activities of SOD, CAT, APX, MDA, soluble protein increased significantly with increased concentration of Se, while the activities of SOD, CAT, and APX increased at 100 μ M Se stress and then CAT activities decreased at 900 μ M stress. POD activities decreased upon exposure to any levels of Se. Se pretreatment had little effect on the enzymatic components of seedlings grown under normal conditions. The results indicate that the exogenous application of Se at low concentrations increases the tolerance of plants to Se-induced oxidative damage by enhancing their antioxidant defense and Se detoxification systems.

For adult human, the recommended dietary allowance of selenium (Se) - an essential micronutrient is estimated at between 40 and 70 μ g per day (World Health Organization) with meat, seafood and cereals being the main sources of dietary Se in China. Selenium is a metalloid that resembles sulphur (S). In trace amounts, Se is considered an essential element for human and animal health (Mehdawi *et al.* 2014). It is a constituent of selenoenzymes such as glutathione peroxidase (GSH-Px), thioredoxin reductases (TR), proteins implicated in Se transport (selenoprotein P) and proteins with unknown functions that are involved in maintaining the cell redox potential (Zeng and Comb 2008, Pazoki *et al.* 2010, Cambrollé *et al.* 2012) because of its critical role in organic antioxidant defense systems (Inostroza-Blancheteau *et al.* 2003) and cancer prevention (Dai *et al.* 2012).

Se is a metalloid that can occur naturally in soils from the Enshi and Ziyang in the western China. Agricultural irrigation and runoff dissolves Se from these soils, causing accumulation of toxic levels of selenate (SeO₄²⁻) in water and soil. Selenate is the most common species of Se found in the root zone (Bañuelos et al. 2010) and can contaminate both water and soil (Mehdawi et al. 20111). Therefore, it has negative effects on plants at high concentrations by reducing growth and causing leaf chlorosis, and nutritional disturbances. The deleterious effects of selenium on photosynthesis is associated with Se-induced decreases in electron transport yields in photosystem II (PSII) and pigment biosynthesis (Germ et al. 2005, Zahedi et al. 2009), as well as Se-induced oxidative damage (Mechora et al. 2013) and (Funes-Collado et al. 2013). In order to detoxify the excess of reactive oxygen species (ROS), plants have antioxidant enzymatic systems involving superoxide dismutase (SOD, EC 1.15.1.1), catalase (CAT, EC 1.11.1.6), ascorbate peroxidase (APX, EC 1.11.1.11), guaiacol peroxidase (POD, EC 1.11.1.7), and glutathione reductase (GR, EC 16.4.2). These functionally interrelated enzymes can provide protection by eliminating ROS to minimize oxidative damage in plant when response to environmental stress. A coordinated increase in enzymatic antioxidants (SOD, APX and GR) was noticed in response to different Se concentrations in ryegrass (Hartikainen et al. 2000, Cartes et al. 2011). Similarly, the antioxidant enzyme activities

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(SOD, CAT and APX) increased in Se-treated sorghum and wheat (Djanaguiraman *et al.* 2010, Inostroza-Blancheteau *et al.* 2013). Redox-active metals (i.e. Zinc) are also known to cause both direct oxidative damage and antioxidative protection (Cambrollé *et al.* 2012), although little is known about the role of non-redox active metals (as such Se) in oxidative pathways.

Alfalfa (*Medicago sativa* L.) is an important leguminous forage and has a wide distribution in the world. In this study, we first constructed Se tolerance and related tolerance mechanisms in alfalfa by evaluating their antioxidant system activities (SOD, CAT, APX and POD) when exposed to increasing Se doses. This study will help us to better understand the physiological regulation mechanisms of forage in response to Se exposure and provide new insights into engineering forage for phytoremediation.

The experiments were performed in the orchard of Shaanxi University of Technology, Hanzhong (33°34'N, 107°28'E), P. R. China. Alfalfa (*Medicago sativa* ssp.) seeds were purchased from Northwest A&F University. Seeds were surface-sterilized with 0.1% (w/v) HgCl₂ for 10 min. Alfalfa (*Medicago sativa* ssp.) seeds were sown in plastic pots filled with a peat/vermiculite matter mix (w/w 1 : 1) and grown in a greenhouse under a day/night temperature of 25/15°C, 500 µmol/m²/s photosynthetic active radiation and a 12 hrs photoperiod. The nutrient solution includes 5 mM Ca(NO₃)₂·4H₂O, 5 mM KNO₃, 2 mM MgSO₄·4H₂O, 1 mM KH₂PO₄, 0.1 mM EDTA-Fe, 461 mM H₃BO₃, 9.11 mM MnCl₂·4H₂O, 0.321 mM CuSO₄·5H₂O, 0.761 mM ZnSO₄.7H₂O and 0.51 mM H₂MoO₄·H₂O (Dai *et al.* 2013). After three-month cultivation, the plants were treated with 0, 100 and 900 µM Se by adding Na₂SeO₃ into the nutrient solution, respectively. Six repeats were obtained for every treatment.

The seedlings were harvested after 20, 40 and 60 days of exposure. The roots were immersed in 5 mM $CaCl_2$ for 15 min and then the whole plants were rinsed with deionized water. The leaves were divided and dried in an air oven at 80°C until a constant dry weight was gained. The effects of zinc treatment on the dry weight of leaf were assessed by two-way ANOVA.

The activity of ascorbate peroxidase (APX, EC 1.11.1.11), superoxide dismutase (SOD, EC 1.15.1.1), catalase (CAT, EC 1.11.1.6) and guaiacol peroxidase (POD, EC 1.11.1.7) after Dai *et al.* (2012) were determined.

Free proline and soluble protein were measured by the method of Dai *et al.* (2015a). The malonaldehyde (MDA) concentrations in plant materials were analyzed spectrophotometrically at 450, 532 and 600 nm as described previously.

A completely randomized design was used for each time point with six replicates. Data were subjected to analysis of variance (ANOVA) to examine the effects of time, treatment and species. Statistical analysis was conducted by using STATISTICA 5.1 software (Statsoft Inc., United States of America). Separation of means was carried out by using Fisher's LSD test at p < 0.01 and p < 0.001 significance levels.

After 20 days of Se exposure, the contents of MDA, soluble protein and free proline significantly increased when the Se stress increased to 100 and 900 μ M, compared to control (non-Se) (Fig.1a,b,c), the MDA concentration in alfalfa leaves increased after 0, 100 and 900 μ M Se exposure, respectively (Fig. 1a). After 40 days, under 100 and 900 μ M Se treatment led to 40.6 and 62.7% increase in MDA levels, and after 60 days, 23.2 and 32.3% enhancement was observed over the untreated control (Fig. 1a). However, after 60 days, free proline content in alfalfa leaves decreased by 4.1% with 100 μ M Se and by 31.6% with 900 μ M Se as compared with control leaves, respectively (p < 0.01). Exposure to 100 and 900 μ M Se also resulted in an increase in the soluble protein content, compared to control (Fig. 1c). After 60 days of Se treatment, the soluble protein level was further increased by 6.3 and 20.3% in comparison with the control, respectively (p < 0.01).

As selenium application increased from 100 and 900 μ M, the activities of SOD and CAT in leaves (Table 1) showed increasing trend during the days 20. After 60 days, SOD activities in alfalfa leaves increased by 3.7% with 100 μ M Se and by 13.4% with 900 μ M Se as compared with control leaves, after 100 μ M Se exposure, the CAT activity in leaves increased 11.3%, however, exposure to 900 μ M Se, the CAT activity was decreased 37.3%, compared to control. After 20 days, APX activity decreased linearly with decreased Se levels, compared to control. In contrast, after 60 days, when compared to the control, the APX activity was increased with the 100 and 900 μ M Se treatment of leaves. According to the results, POD activity decreased linearly with increasing Se levels. The highest concentration of Se (900 μ M) proved to be extremely toxic resulting in a decline of POD activity (Table 1).



Fig. 1. Changes of MDA, free proline and soluble protein contents in alfalfa leaves under Se stress. Values are means of six replicates \pm standard deviation. Different small alphabets are representing the significant difference at $p \le 0.01\%$.

Plant stress tolerance to heavy metals is often correlated to an increase in the activity of antioxidant enzymes (Dai *et al.* 2012, Dai *et al.* 2015b). Plants which do not have strong antioxidative enzymes system may experience oxidative stress. In addition to nonenzymatic antioxidants, antioxidative enzymes are of great importance for the plants against ROS, such as APX, SOD and CAT (Pazoki *et al.* 2010, Freeman and Bañuelos 2011). The increased activity

Se (µM)	Time (days)	SOD	CAT	POD	APX
Control	20	$307.7\pm30.1b$	$21.2 \pm 2.1d$	$35.3 \pm 3.9b$	$257.1 \pm 22.2c$
	40	$252.8\pm23.8d$	$48.4\pm4.6bc$	$47.4 \pm 4.1a$	$385.7\pm32.6b$
	60	$257.5\pm23.3cd$	$56.5 \pm 5.1b$	$37.6\pm3.2b$	$128.6 \pm 11.2e$
	20	$370.2\pm29.1a$	$35.3 \pm 3.3c$	$37.1 \pm 3.1b$	$118.6 \pm 10.2e$
100	40	$265.1 \pm 26.6c$	$42.4 \pm 4.1 bc$	$51.8 \pm 5.1a$	$259.2 \pm 23.5c$
	60	$267.1 \pm 23.6c$	$50.1\pm4.8b$	$34.3 \pm 3.5 bc$	$158.6\pm13.8d$
	20	$332.3\pm31.2ab$	$49.4\pm5.0ab$	$28.2 \pm 2.9c$	$214.3\pm19.6cd$
900	40	$273.3\pm28.1c$	$54.5\pm5.3b$	$32.9 \pm 3.1c$	$514.3 \pm 49.2a$
	60	$292.0\pm28.3b$	$77.6 \pm 7.2a$	$25.9\pm2.2d$	$200.6\pm18.8d$
p values	Se	**	***	***	**
	Time	***	**	**	***
	$Se \times time$	***	***	ns	***

Table 1. Changes of SOD, CAT, APX and POD activities (U/mg FW) of alfalfa leaves grown with 0 (control), 100 and 900 µM Se for 20, 40 and 60 days.

Each point represents the mean of six replicates \pm S.E. Data were presented as mean \pm St (n = 6). Different letters behind the values in the same column indicate significant difference between the treatments, p values of ANOVA of Na₂SeO₃ (Se), time and their interactions were also shown. Represents the significance level at *p $\leq 0.05\%$, **p $\leq 0.01\%$; *** p $\leq 0.001\%$. ns, not significant.

of antioxidative enzymes in a plant indicates the formation of ROS, and the (Dai *et al.* 2015a, Zhao and Dai *et al.* 2015) indicated the generation of oxidative stress in plants since all enzyme activities increased with high Se levels, which was consistent with our experimental results. Here, we showed that alfalfa responded to Se exposure with increases in concentrations of soluble protein in leaves and with enzymatic activities of APX, SOD and CAT in leaves (Table 1). These results indicated that the antioxidant system is enhanced and well-orchestrated in the leaves of alfalfas under different Se concentrations.

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References

- Bañuelos GS, Stushnoff C, Walse SS, Zulen T, Yang SI and Pickering I 2012. Biofortified, selenium enriched fruit and cladode from three *Opuntia* cactus pear cultivars grown on agricultural drainage sediment for use in nutraceutical foods. Food. Chem. 135: 9-16
- Cambrollé J, Mancilla-Leytón JM, Muñoz-Vallés S, Luque T and Figueroa ME 2012. Zinc tolerance and accumulation in the salt-marsh shrub *Halimione portulacoides*. Chemosphere **86**: 867-874.
- Cartes P, Gianfreda L, Paredes C and Mora ML 2011. Selenium uptake and its antioxidant role in ryegrass cultivars as affected by selenite seed pelletization. J. Soil. Sci. Plant. Nutr. **11**: 1-14

- Dai HP, Shan CJ, Lu C, Jia GL, Wei AZ, Sa WQ and Yang TX 2012b. Response of antioxidant enzymes in populus × canescens under cadmium stress. Pak. J. Bot. 44(6): 1943-1949.
- Dai HP, Shan CJ, Hua Z, Li JC, Jia GL, Jiang H, Wu SQ and Wang Q 2015a. The difference in antioxidant capacity of four alfalfa species in response to Zn. Ecotox. Environ. Saf. **114**: 312-317.
- Dai HP, Jia GL and Shan CJ 2015. Jasmonic acid-induced hydrogen peroxide activates MEK1/2 in upregulating the redox states of ascorbate and glutathione in wheat leaves. Acta. Physiol. Plant. 37: 200-206.
- Djanaguiraman M, Prasad PVV and Seppanen M 2010. Selenium protects sorghum leaves from oxidative damage under high temperature stress by enhancing antioxidant defense system. Plant. Physiol. Biochem. 48: 999-1007.
- Freeman JL and Bañuelos GS 2011. Selection of salt and boron tolerant selenium hyperaccumulator *Stanleya pinnata* genotypes and characterization of Se phytoremediation from agricultural drainage sediments. Environ. Sci. Technol. **45**: 9703-9710.
- Funes-Collado V, Morell-Garcia A, Rubio R and López-Sánchez JF 2013. Selenium uptake by edible plants from enriched peat. Sci. Horticul. **164**: 428-433.
- Germ M, Kreft J and Osvald J 2005. Influence of UV-B exclusion and selenium treatment on photochemical efficiency photosystem, yield and respiratory potential in pumpkins (*Cucurbita pepo* L.). Plant. Physiol. Biochem. 43: 445-448.
- Hartikainen H, Xue T and Piironen V 2000. Selenium as an antioxidant and pro-oxidant in ryegrass. Plant Soil. 225:193-200.
- Inostroza-Blancheteau C, Reyes-Díaz M, Alberdi M, Godoy K, Rojas-Lillo Y, Cartes P and Mora M L 2013. Influence of selenite on selenium uptake, differential antioxidant performance and gene expression of sulfate transporters in wheat genotypes. Plant. Soil 369: 47-59.
- Mehdawi EAF, Quinn CF and Pilon-Smits EAH 2011. *Selenium hyperaccumulators* facilitate selenium-tolerant neighbors via phytoenrichment and reduced herbivory. Curr. Biol. **21**: 1440-1449.
- Mehdawi EAF, Cappa JJ, Fakra SC, Self J and Pilon-Smits EAH 2012. Interactions of selenium and non-accumulators during co-cultivation on seleniferous or non-seleniferous soil - The importance of having good neighbors. New. Phytol. 194: 264-277
- Mehdawi AFE, Reynolds RJB, Prins CN, Lindblom SD, Cappa J J, Fakra SC and Pilon-Smits EAH. 2014. Analysis of selenium accumulation, speciation and tolerance of potential selenium hyperaccumulator Symphyotrichum ericoids. Physiol. Plant. 152: 70-83.
- Mechora S, Stibilj V and Germ M 2013. The uptake and distribution of selenium in three aquatic plants grown in Se(IV) solution. Aquat. Toxicol. 128-129: 53-59.
- Pazoki AR, Shirani-Rad AH, Habibi D, Paknejad F, Kobraee S and Hadyat N 2010.Effect of drought stress and selenium spraying superoxide dismotase activity of winter rapeseed (*Brassica nnapus* L) cultivars. World. Acad. Sci. Eng. Technol. 68: 678-681.
- Zahedi G, Noormohammadi G, Shirani-Rad AH, Habibi D and Boojar MMA 2009. Effect of zeolite and foliar application of selenium on growth, yield and yield component of three canola cultivar under drought stress. World. Appl. Sci. J. **7**: 255-262.
- Zeng H and Combs Jr GF 2008. Review. Selenium as an anticancer nutrient: roles in cell proliferation and tumor cell invasion. J. Nutr. Biochem. **19**: 1-7.
- Zhao H and Dai HP 2015. Physiological Response of *Apocynum Venetum* seedings under osmotic stress. Bangladesh J. Bot. **44**(4): 551-556.

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